Benzimidazoles: A New Class of Carbonic Anhydrase Inhibitors

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Summary: Carbonic anhydrase inhibitory activity of benzimidazole derivatives **1-24** has been evaluated. Compounds **22** (IC₅₀ = 7.47 ± 0.39 μ M), **21** (IC₅₀ = 10.31 ± 0.11 μ M), **20** (IC₅₀ = 23.1 ± 1.78 μ M), **12** (IC₅₀ = 12.16 ± 0.10 μ M), **11** (IC₅₀ = 12. 81 ± 0.29 μ M), and **15** (IC₅₀ = 13.18 ± 0.37 μ M) showed carbonic anhydrase inhibitory activity, and compared with the standard acetozolamide (IC₅₀ = 0.31 ± 0.0006 μ M). All the compounds were characterized by spectroscopic techniques.

Keywords: Benzimidazole, carbonic anhydrase inhibition, phenylenediamine, aromatic aldehydes

Introduction

In recent years modified benzimidazoles are reported to exhibit a broad spectrum of biological activities including antifungal and anticancer activities [1, 2]. Benzimidazole derivatives have also been found to possess various biological activities such as antibacterial [3-7], antihelmintic [8, 9], epileptic, antdiabetic, anti-fertility [10, 11], antiviral [12-15], antiinflammatory [16], antihistaminic [17], antioxidant [18-21], antihypertensive [22], anticoagulant [23], proton pump inhibitory [24, 25], antileukaemic [26], and antiulcer properties.

zinc Previously benzimidazole-based complexes were used as structural carbonic anhydrase models. Furthermore, benzimidazole and indapamide-like benzenesulfonamides have been reported as Rho kinase and carbonic anhydrases I, II, VII, and VIII, respectively [27]. We present here the in vitro carbonic anhydrase inhibitory activity of a series of benzimidazole derivatives 1-24. Carbonic anhydrase (EC 4.2.1.1) is a Zn²⁺ containing enzyme and composed of a single polypeptide bonds, and a tightly bound Zn^{2+} ion is required for activity of this enzyme, it catalyzes the interconversion of carbon dioxide and water to bicarbonate and proton and thus maintain acid-base balance in blood and other tissues. It plays a key role in transportation of carbon dioxide out of the tissues. These are the known physiological function of carbonic anhydrase; it also affects on the diverse processes, such as physiological pH control, gas balance, and calcification. Since last three decades carbonic anhydrase inhibitors have been used in the treatment of glaucoma. Sulfonamide, a carbonic anhydrase inhibitor, has been used in the

treatment of glaucoma [28], whilst, acetazolamide is found to potent carbonic anhydrase inhibitor, useful in the management of glaucoma [29-32], which opened new avenues for the medicinal chemists to work on other carbonic anhydrase inhibitors. Currently used carbonic anhydrase inhibitors are administered systemically, include acetazolamide, dichlorophenamide, ethoxzolamide and methazolamide. Each of these drugs possesses a free sulfamoyl group, attached to an aromatic heterocyclic ring and inhibits the enzyme *in vitro* with potency in the nanomolar range [31]. Intraocular pressure is decreased by reduction in humor formation stemming from the inhibition of carbonic anhydrase, present in the ciliary epithelium. A number of carbonic anhydrase inhibitors have been reported to lower the intraocular pressure when instilled topically in animals [33]. In the present study, benzimidazoles were randomly screened for in vitro inhibition of bovine carbonic anhydrase. The carbonic anhydrase inhibitory activity exhibited by compound 22 is 24 fold lower than the standard (IC_{50} = 0.31 \pm 0.0006 μ M) Table-1. This represents the potential of this class of compounds as carbonic anhydrase inhibitors for possible treatment of associated diseases.

Chemistry

Benzimidazoles 1-24 were synthesized by reacting together with commercially available 2phenylenediamine and different aromatic aldehydes in N,N-dimethylformamide (DMF). The products were appeared in significant yields (Scheme-1). In a typical reaction, sodium metabisulfite (Na₂S₂O₅) was added to the stirring mixture of 2-phenylenediamine

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(3.12 mmol) and different substituted aromatic aldehydes (3.16 mmol) in *N*,*N*-dimethylformamide. Reaction mixture was refluxed for 2 h. The progress of reaction was monitored by TLC and after completion of reaction; mixture was allowed to cool to room temperature. Water (30 mL) was added to the crude reaction mixture to afford solid mass as precipitates, after filtration the solid benzimidazole derivatives **1-24** were obtained in high yields. Recrystallization from methanol afforded pure products and the structures of synthetic compounds **1-24** were evaluated by spectroscopic techniques including ¹H-NMR, and EI MS [34].

		NH2 NH R1	+ 0 R ₂	DM	$_{2}S_{2}O_{5}$ F, Reflux 2h	N N R ₁			
Comp. No.	R ₁	R ₂	Comp. No.	R ₁	R ₂	Comp. No.	R ₁	R ₂	
1	Н	⁶ 5' OMe OMe	9	Н	6' F 5' 4' 3'	17	Н	6' 2' 5' 3'	
	Н	6' OH 5' OH OH	10	Н	2' 3' 4' 5' 6'	18	Н	6 1' 2' 5' F 3'	
	Н	6 5 0 0 H	11	н	⁶ 5' OH 4' OMe	19	Ph		
	Н	1' 8' 7' 3' 4' 5' 6'	12	Н	6 5' 3'	20	Ph	2' 3' 4' 5' 6'	
	Н	6 5'2' 4' OH	13	н	6' Me	21	Ph	$ \begin{array}{c} 6' \\ 5' \\ 4' \end{array} $ Me	
	Н	$ \begin{array}{c} 9' \\ 8' \\ 7' \\ 6' \\ 5' \\ 5' \end{array} $	14	Н	$\begin{array}{c} 6' \\ 5' \\ OEt \end{array}$	22	Ph	ome OMe	
	Н	HO 4' S'	15	Н	6'	23	н	6 Cl 4'	
	Н	6' 2' 5' 4' Cl	16	Н	6 5' OH OEt	24	н	6 5' OH OH	
Scheme-1	:	Synthetic benzimida	zole derivati	ves 1-					
Table-1: In vitro carbonic anhydrase activity of compounds 1-24.									

S. No.	% Inhibition	$IC_{50} \pm SEM^{a}(\mu/M)$	S. No.	% Inhibition	$IC_{50} \pm SEM^{a} (\mu/M)$
1	15.9	NA ^b	13	5.9	NA ^b
2	17.1	NA^{b}	14	- 4.9	NA ^b
3	13.5	NA^{b}	15	62.2	13.18 ± 0.37
4	Precipitated	NA^{b}	16	Precipitated	NA ^b
5	29.1	NA^{b}	17	6.4	NA ^b
6	Precipitated	NA^{b}	18	40.9	NA ^b
7	41.2	NA^{b}	19	Precipitated	NA ^b
8	21.3	NA^{b}	20	62.3	23.1 ± 1.78
9	26.3	NA ^b	21	83.1	10.31 ± 0.11
10	21.4	NA^{b}	22	79.7	7.47 ± 0.39
11	61.1	12.81 ± 0.29	23	34.8	NA ^b
12	78.2	12.16 ± 0.10	24	- 3.2	NA ^b
Acetozolamide (Standard)		0.31 ± 0.002			

SEM^a: is the standard error of the mean, NA^b: Not active Acetozolamide; standard inhibitor of Carbonic anhydrase.

Bioactivity

Twenty four (24) synthetic benzimidazole derivatives were screened for their carbonic anhydrase inhibitory potential. Six of them showed above 50% inhibition, we therefore screened them for their IC₅₀ values. Compounds **22** (IC₅₀ = 7.47 ± 0.39 μ M), **21** (IC₅₀ = 10.31 ± 0.11 μ M), **20** (IC₅₀ = 23.1 ± 1.78 μ M), **12** (IC₅₀ = 12.16 ± 0.10 μ M), **11** (IC₅₀ = 12. 81 ± 0.29 μ M), and **15** (IC₅₀ = 13.18 ± 0.37 μ M) showed good carbonic anhydrase inhibitory activity. Acetozolamide, (IC₅₀ = 0.31 ± 0.0006 μ M) was used as standard in the assay.

Limited SAR studies revealed that the carbonic anhydrase inhibitory activity is dependent on the nature of R_1 and R_2 substitutions. The highest inhibition was demonstrated by the compound 22 $(IC_{50} = 7.47 \pm 0.39 \ \mu M)$, which has a phenyl group as R_1 and 3',4'-dimethoxy phenyl as R_2 on benzimidazole skeleton. Interestingly, compound 1 was found to be completely inactive, though it has same R₂ substituents on benzimidazole ring. The comparison of activity of compounds 1 and 22 indicate that the activity is largely due to R_1 phenyl substituents. Compounds 20-22 contain R₁ as phenyl substituents have shown diverse level of activities. Compound **21** showed an IC₅₀ value $10.31 \pm 0.11 \ \mu M$ as the second most active compound of the series. Likewise its analogous compound 13 with same R₂ showed inactivity. Compound 12 contain an isopropyl group at phenyl as R2 and H as R1 showed an activity with an IC₅₀ value $10.31 \pm 0.11 \ \mu$ M. Compound **11** with an IC₅₀ value 12. $81 \pm 0.29 \ \mu$ M was found to be fourth most active compound of the series which contains 2-hydroxy and 3-methoxy at phenyl as R₂. Compound 15 with an IC₅₀ value 13.18 \pm 0.37 μ /M contains 3,4-dichloro phenyl as R₂. In addition, compound **20** (IC₅₀ = $23.1 \pm 1.78 \mu$ M) having a 1-naphthyl as R_2 and phenyl as R_1 but its analogous compound 10 found to be completely inactive. This further indicates the importance of R₁ as phenyl as well right substituents on R₂. This SAR is by no means conclusive and needs evaluation of a larger library of benzimidazoles.

This study, however, demonstrates that compound **22** can be an initial point for further study hence its chemical modification may lead to more potent carbonic anhydrase inhibitors.

Material and Methods:

The carbonic anhydrase was purchased from Sigma Aldrich (Cat. No. C-3934), 4-nitrophenyl acetate was purchased from MP Bio (Cat. No. 160888). HEPES buffer was purchase from DOJINDO (Cat. No. 346-01373), *tris*-(hydroxymethyl)-aminomethane (reagent grade) was purchase from Scharlau (Cat. No. TR0423). DMSO and ethanol of reagent grade were used in the experiment. The experiment was performed in triplicate. IC_{50} values were calculated by using EZ-fit enzyme kinetics software (USA)

Carbonic Anhydrase Inhibition Assay

In this assay, colorless 4-nitrophenyl acetate (4-NPA), is hydrolyzed to 4-nitrophenol and CO_2 and reaction is followed by measuring the formation of 4-nitrophenol, a yellow colored compound.

The experiment was performed in buffer containing HEPES and *Tris* at a total concentration of 20 mM and pH was 7.4. For each sample the reaction tube contained 140 μ L of the *HEPES-Tris* solution, 20 μ L of freshly prepared aqueous solution of purified bovine erythrocyte carbonic anhydrase II (0.1 mg/mL of deionized water for 96-well), 20 μ L of test compound dissolve in DMSO (10% final concentration), 20 μ L of substrate 4-PNA at concentration of 0.7 mM diluted in ethanol.

The reaction was initiated by addition of 4-PNA after 15 min incubation of test compounds. The compounds were tested in triplicate at different concentration. In this assay the reaction was performed in 96-well plate. *SPECTRA-max* 340 spectrophotometer, Molecular Devices (USA) was used to monitor the reaction and the amount of product formed was monitored at 1 min interval for 30 min at 400 nm [35, 36].

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References

- 1. S. A. Wank, American Journal of Physiology-Gastrointestinal and Liver, **274**, 607 (1998).
- 2. H. M. Refaat, *European Journal of Medicinal Chemistry*, **45**, 2949 (2010).
- C. Hubschwerlen, P. Pflieger, J. L. Specklin, K. Gubernator, H. Gmünder and P. I. A, Kompis, *Journal of Medicinal Chemistry*, 35, 1385 (1992).
- 4. H. Göker, M. Tunçbilek, G. Ayhan and N. Altanlar, *IL Farmaco*, **53**, 415 (1998).

Uncorrected Proof

- P. T. M. Nguyen, J. Baldeck, J. Olsson and R. E., Marquis, *Oral Microbiology and Immunology*, 20, 93 (2005).
- S. Ozden, D. Atabey, S. Yıldız and H. Göker, Bioorganic and Medicinal Chemistry, 13, 1587 (2005).
- H. Goker, S. Ozden, S. Yıldız amd D. W. Boykin, *European Journal of Medicinal Chemistry*, 40, 1062 (2005).
- 8. L. Veerakumari and N. Munuswamy, *Veterinary Parasitology*, **91**, 129 (2000).
- G. Merino, J. W. Jonker, E. Wagenaar, M. M. Pulido, A. J. Molina, A. I. Alvarez and A. H. Schinkel, *Drug Metabolism and Disposition*, 33, 614 (2005).
- A. Orjales, R. Mosquera, L. Labeaga and R. Rodes, *Journal of Medicinal Chemistry*, 40, 586 (1997).
- M. Grimmett, Comprehensive Heterocyclic Chemistry: The Structure, Reactions, Synthesis and Uses of Heterocyclic Compounds vol. 3, Pergamon, Oxford, p. 77 (1996).
- S. Budow, M. Kozlowska, A. Gorska, Z. Kazimierczuk, H. Eickmeier, P. L. Colla, G. Gosselin, F. Seela, *Arkivoc*, 3, 225 (2009).
- T. Ishida, T. Suzuki, S. Hirashima, K. Mizutani, A. Yoshida, I. Ando, S. Ikeda, T. Adachi and H. Hashimoto, *Bioorganic and Medicinal Chemistry Letters*, 16, 1859 (2006).
- 14. L. Garuti, M. Roberti and G. Gentilomi, *IL Farmaco*, **56**, 815 (2001).
- K. K. Biron, R. J. Harvey, S. J. Chamberlain, S. S. Godd, A. A. Smith III, M. G. Davis, C. L. Talarico, W. H. Miller, R. Rerris, R. E. Dornsife, S. C. Stanat, J. C. Drach, L. B. Townsend and G. W. Koszalka, *Antimicrobial Agents and Chemotherapy*, 46, 2365 (2002).
- K. C. S. Achar, K. M. Hosamani and H. R. Seetharamareddy, *European Journal of Medicinal Chemistry*, 45, 2048 (2010).
- R. Iemura, T. Kawashima, T. Fukuda, K.. Ito and G. Tsukamoto, *Journal of Medicinal Chemistry*, 29, 1178 (1986).
- E. R. Cole, G. Crank and S, A Salam, *Journal of Agricultural and Food Chemistry*, 22, 918 (1974).
- C. Kus, G. Ayhan-KIlcIgil, B. C. Eke and M. Iscan, *Archives of Pharmacal Research*, 27, 156 (2004).
- O. Temiz-Arpact, T. Coban, B. Tekiner-Gulbas, B. Can-Eke, I. Yildiz, E. Aki-Sener, I. Yalcin and M. Iscan, *Acta Biologica Hungarica*, 57, 201 (2006).

- Z. Ates-Alagoz, C. Kus and T. Coban, *Journal of Enzyme Inhibition and Medicinal Chemistry*, 20, 325 (2005).
- 22. J. R. Kumar, J. L. Jat and D. P. Pathak, *E-Journal of Chemistry*, **3**, 278 (2006).
- Z. Zhao, D. O.Arnaiz, B. Griedel, S. Sakata, J. L. Dallas, M. Whitlow, L. Trinh, J. Post, A. Liang, M. M. Morrissey and K. J. Shaw, *Bioorganic and Medicinal Chemistry Letters*, 10, 963 (2000).
- T. C. Kuhler, M. Swanson, V. Shcherbuchin, H. Larsson, B. Mellgard and J. E. Sjostrom, *Journal* of *Medicinal Chemistry*, 41, 1777 (1998).
- 25. J. Horn, Clinical Therapeutics, 22, 266 (2000).
- 26. a) A. Kamal, P. P. Kumar, K Sreekanth, B. N. Seshadri and P. Ramulu *Bioorganic and Medicinal Chemistry Letters*, 18, 2594 (2008). b) L. Garuti, M. Roberti, M. Malagoli, T. Rossi, M. Castelli, *Bioorganic and Medicinal Chemistry Letters*, 10, 2193 (2000).
- 27. a) M. M Ibrahim, M. A Amin and K. Ichikawa, Journal of Molecular Structure, 895, 191 (2011).; b) E. Capkauskaite, L. Baranauskiene, D. Golovenko, E. Manakova, S. Grazulis, S. Tumkevicius and D. Matulis, Bioorganic and Medicinal Chemistry, 13, 1587 (2005).; c) Feng, Y.; Lograsso, P.; Bannister, T.; Schroeter, T.; Fang, X.; Chen, Y. T.; Yin, Y.; Smolinski, M. P.; Yao, L.; Wang, B.; Sessions, H. (2009) (#WO2009079009).
- T. H. Maren, Drug Development Research, 10, 255 (1987).
- A. Bar-Iian, N. I. Pessah and T. H. Maren, Investigative Ophthalmology and Visual Science, 25, 1198 (1984).
- 30. E. B. Werner, D. S. Gerber and J. Y. Yolando, *Journal of Ophthalmology*, **22**, 316 (1987).
- M. F. Sugrue, P. Gautheron and C. Schmitt, Journal of Pharmacology and Experimental Therapeutics, 232, 534 (1985).
- R. A. Lewis, R .Schoenwald, C. F. Barfknecht and C. D. Phelps, *Archives of Ophthalmology*, 104, 842 (1986).
- 33. J. E. A. McIntosh, *Biochemical Journal*, **120**, 299 (1970).
- K. M. Khan, M. Khan, N. Ambreen, F. Rahim, S. Naureen, S Perveen,; M. I. Choudhary, W. Voelter, *Medicinal Chemistry*, 8, 421 (2012).
- P. S. Richard, R. D. Dennis, J. S. Anthony and B. Meir, *Epilepsy Research*, 63, 103 (2005).
- 36. O. Arsalan, Biochemistry, 66, 982 (2001).