

Bisbenzylisoquinoline Alkaloids from *Epinetrum villosum* (Menispermaceae)M. PARVEZ, ABDUR RAHMAN*, SALMA RAHMAN AND O.N. OGBEIDE
Institute of Chemistry, University of the Punjab, Lahore, Pakistan

(Received July 22, 1993, revised April 4, 1994)

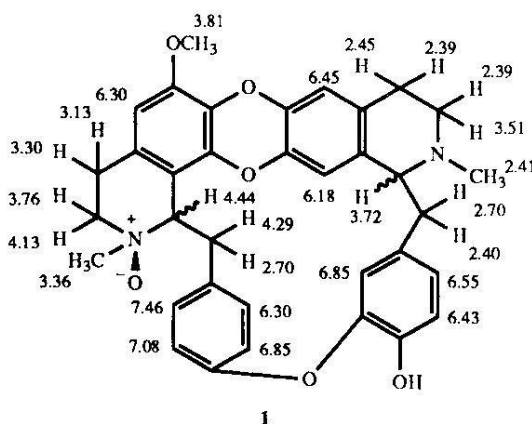
Summary: A phytochemical investigation of the stem bark of *Epinetrum villosum* (Menispermaceae) resulted in the isolation of six bisbenzylisoquinoline alkaloids: cycleanine, cycleanine *N*-oxide, norcycleanine, cocsoline, cocsuline, and a new alkaloid cocsuline *N*-oxide.**Introduction**

Epinetrum villosum (Exell) Troupin (*Albertisia villosa* Farn), a woody climber of Triclisieae tribe of the Menispermaceae family is found in secondary forests in Xerophytic woods on hills in the coastal area of Zaire and Angola. The plant is used natively for the treatment of pains and mental strain [1,2,3]. In 1958, Denoil [4] and in 1971, Bouquet and Cave [5] reported the isolation of cycleanine, norcycleanine and isochondodrine from this plant. We now report in addition to these alkaloids, the isolation of cycleanine *N*-oxide, cocsoline and a new alkaloid cocsuline *N*-oxide. These alkaloids have been identified by means of spectral data (UV, MS, ¹H-NMR) TLC and colour reactions [5-22].

Results and Discussion

The five known alkaloids: cycleanine, cycleanine *N*-oxide, norcycleanine, cocsoline and cocsuline were identified by direct comparison of UV; IR; TLC; mp; ¹H-NMR and colour reactions with authentic samples [5-14]. The new alkaloid proposed to be cocsuline *N*-oxide gave mp. 210°-212° C; [α]_D²⁵ + 264° (C = 0.3, CHCl₃); λ_{max} (CH₃OH) nm. 278, 291, 305 (similar to that of cocsuline), [13,14]; ν_{max} (KBr) similar to that of cocsuline except for an additional peak at 1500 cm⁻¹ which is indicative of N-O stretch of *N*-oxide [21,22]. The mass spectrum m/z 578 (M⁺, 95%) corresponding to C₃₅H₃₄N₂O₆, (16 mass units higher than that of cocsuline C₃₅H₃₄N₂O₅), and a prominent peak m/z 562 (M⁺ - 16, 45%) due to the loss of an oxygen atom suggests the presence of an *N*-oxide function [15,16,23]. The ¹H-NMR spectrum was similar to that of cocsuline except that one of the *N*-methyl signal was shifted down field (δ 3.36 as opposed to δ 2.56) suggesting again the presence of *N*-oxide function [17,18].

We therefore, report that the new alkaloid is an *N*-oxide of cocsoline and has the possible stereo structure 1 (Fig. 1). Further substantiation was done by, reduction of 1 with 10% H₂SO₃ which afforded cocsuline, identical by a direct comparison (UV, IR, TLC properties and chromogenesis) with an authentic sample of cocsuline obtained from *Synclisia scabrida* [12], and oxidation of the authentic sample of cocsuline obtained from [12], with 15% H₂O₂ to get cocsuline *N*-oxide which showed TLC and colour reactions identical to alkaloid 1. These observations confirmed that the new alkaloid 1 is *N*-oxide of cocsoline. It is unlikely that the alkaloid is an artifact formed during the isolation procedure as it can be detected by TLC in the test extracts of *Epinetrum villosum* stem bark.

Fig. 1: Possible structure of cocsuline *N*-oxide**Experimental***Plant material*

The plant material was obtained from Mr. Breyné Heiman (39A rue van Page, 1080,

*To whom all correspondence should be addressed.

Bruselle), Herbarium I.N.E.R.A., Kinshasa, Zaire. A voucher specimen had been deposited at the Department of Chemistry, Bendel State University, Ekpoma.

Extraction, chromatography and detection

The bark was removed from the stems of *Epinetrum villosum*, dried and ground to a coarse powder. The pulverised stem bark (1 Kg) was extracted with 90% MeOH (3 dm³) in a glass percolator to yield a syrupy residue (5 g) after removal of solvent in vacuo. The residue was dissolved in 300 cm³ of 3% HCl (pH-2). After basification with 25% NH₄OH (pH-8), the solution was extracted with CHCl₃ (3 x 150 cm³). The CHCl₃ extracts were combined, washed with H₂O, dried over anhydrous Na₂SO₄ and evaporated to dryness in vacuo, after filtration, to yield a non-quaternary alkaloid fraction (2.0 g).

The nonquaternary alkaloid fraction was dissolved in CHCl₃ (50 cm³) and chromatographed over a silica gel (type 60, 100 mesh ASTM) column (3.5 x 40 cm) packed in CHCl₃. Elution was begun with CHCl₃ and the polarity was increased gradually by addition of MeOH (10%, 15% and 25%). Fraction of 500 cm³, were collected and monitored by TLC. Eventual elution with CHCl₃ - MeOH (4:1) afforded a fraction (1.0 g) containing bisbenzyliso-quinoline alkaloids with two or three diphenyl ether linkages. Final separation was achieved by means of repeated preparative TLC, on 20 x 20 cm, precoated glass plates with 0.5 mm thick layer of silica gel 60 F₂₅₄ (Merck) in the solvent systems: CHCl₃ - MeOH - conc. NH₄OH (10:2:0.1) and EtOAc-(Me)₂CHOH 25% NH₄OH (9:7:2). The second solvent system was used to determine the R_f values of the separated alkaloids.

The alkaloidal zones were detected under UV₂₅₄, UV₃₆₆ and iodoplatinate reagent. Two colour reactions were performed by means of (i) 0.2 M FeCl₃ in 35% HClO₄ and (ii) a new colour "colour ring test", to distinguish the bisbenzylisoquinolines with two and three ether linkages. A developed chromatogram after spraying with H₂PtCl₆ - KI (1:10), dried at 25°C and again sprayed with HNO₃ - HCl - H₂SO₄ (1:1:1) and heated at 100°C for exact 5 min. A bisbenzylisoquinoline with two diphenyl ether linkages (cycleanine) gave a reddish brown spot

surrounded by a yellow, grey and violet rings, while the one with three diphenyl ether linkages (cocsuine) gave a reddish spot surrounded by a violet ring.

Identification

UV spectra were recorded in methanol on a Perkin Elmer model 137UV instrument. The IR spectra were taken as KBr disc on a Perkin Elmer model 257 grating infrared spectrophotometer. Optical rotations were registered with a Perkin Elmer model 141 polarimeter (Na-590 nm). Melting points (uncorrected) were determined using a hot-stage microscope. ¹H-NMR spectra were obtained in CDCl₃ on a Hitachi Perkin Elmer model R-24, high resolution spectrometer.

Cycleanine

¹H-NMR: 2.60 (2 x N-CH₃), 3.40 (2 x O-CH₃), 3.82 (2 x O-CH₃) 4.31 (d, J=10-Hz, 2H), 5.81 (dd, J=8.5 Hz, J=2.7Hz, 2H), 7.1 (dd, J=8.5 Hz, J=2.0 Hz, 2H), (ArH), R_f 0.75; [α]_D²⁵ - 18.2° (C = 0.30, MeOH), UV: 275, 285 nm. Colour reaction with FeCl₃ reagent gave an intense purple colour in hot air.

The data is identical with those for cycleanine [6-9].

Cycleanine N-oxide

¹H-NMR: 2.60 (N-CH₃), 3.32 (N'-CH₃), 3.42 (2 x O-CH₃), 3.84 (2 x O-CH₃), 4.31 (d, J=10Hz, 2H), 5.81 (dd, J = 8.5 Hz, J = 2.7 Hz, 2H), 6.27 (dd, J = 8.5 Hz, J = 2.0 Hz, 2H) 6.58 (dd, J = 8.5Hz, J = 2.7 Hz, 2H), 7.1 (dd, J=8.5, J=2.0Hz, 2H) (ArH). R_f 0.37; [α]_D²⁵ - 7.6° (C=0.30, MeOH). UV, 275, 285 nm. Colour reaction with FeCl₃ reagent gave an intense purple colour to hot air.

The data is identical with those for cycloeanine N-oxide [12].

Norcycleanine

¹H-NMR: 2.6 (2 x N-CH₃), 3.40 (O-CH₃), 3.81 (O-CH₃), 3.84 (O-CH₃) 4.31 (d, J=10 Hz, 2H), 5.79 (bd, J=8.5 Hz, 2H), 6.25 (d, J = 8.5 Hz, 3H), 6.50 (s, 1H), 6.56 (s, 1H), 6.50 - 6.66 (m, 2H), 6.97 - 7.12 (m, 2H) (ArH). R_f 0.65 [α]_D²⁵ + 23.5° (C=0.08, MeOH), UV: 275, 285 nm. Colour reaction with FeCl₃ reagent gave a brownish purple colour in hot air.

The data is identical with those for norcycleanine [7,10,11].

Cocsuline

¹H-NMR: 2.60 (N-CH₃), 3.85 (O-CH₃), 6.19 (s, 1H), 6.33 (s, 1H), 6.54 (d, J=2.7 Hz, 2H), 6.62 (s, 1H), 6.76-7.22 (m, 4H), 7.46 (dd, J = 8Hz, J = 2.7 Hz, 1H), (ArH). R_f 0.57; [α]_D²⁵ + 198° (C = 0.08, CHCl₃), UV: 275, 285, 302 nm. Colour reaction with FeCl₃ reagent gave a greenish black colour in hot air.

The data is identical with those for cocsuline [13,14].

Cocsuline

¹H-NMR 2.43 (N-CH₃), 2.56 (N-CH₃), 3.81 (O-CH₃), 3.76 (s, 1H), 4.13 (s, 1H), 6.18 (s, 1H), 6.30 (s, 1H), 6.45 (s, 1H), 6.55 (d, J = 2.7Hz, 2H), 6.85 (s, 1H), 6.93 (dd, J = 8.0Hz, J = 2.7 Hz, 2H), 7.08 (bm, 1H), 7.46 (bd, J=8.0 Hz, 1H) (ArH). R_f 0.63; [α]_D²⁵ + 275° (C = 0.30, CHCl₃). UV: 278, 291, 305 nm. Colour reaction with FeCl₃ reagent gave a greenish-black colour in hot air.

The data is identical with those for cocsuline [13,14].

Cocsuline N-oxide

¹H-NMR: 2.40 dd, 2.41 (N-CH₃), 2.45 m, 2.70 dd, 2.93 m, 3.13 m, 3.30 m, 3.36 (N-CH₃), 3.51 m, 3.72 brd, 3.76 m, 3.81 (O-CH₃), 4.13 m, 4.29 (dd, 4.8, 12.3 Hz), 4.44 dd, 6.18 (s, 1H), 6.30 (s, 1H), 6.45 (s, 1H), 6.55 (dd, J = 2.7 Hz 2H), 6.85 (s, 1H), 6.93 (dd, J = 8.0, 2.7 Hz, 2H), 7.08 (bm, 1H), 7.46 (bd, J=8.0 Hz, 1H) (ArH). MS (380°C, 70eV): m/z 578 (M⁺, 95), 562 (M⁺-16, 45). R_f 0.35; m.p. 210° - 212°C. UV: 278, 291, 305 nm. ν_{max} (KBr) cm⁻¹ 3400, 2910, 1610, 1500, 1220, 1110, 1020, 830. [α]_D²⁵ + 264 (C = 0.30, CHCl₃).

Colour reaction with FeCl₃ reagent gave a greenish black colour in hot air and with a mixture of conc. H₂SO₄ + HNO₃ (1:1) a blue colour indicative of an alkaloid with a dibenzo-1, 4-dioxin moiety [19,20]. In colour ring test, a reddish spot surrounded by a violet ring. These colour reactions were similar to that of cocsuline.

Reduction of cocsuline N-oxide

0.5 mg of the cocsuline N-oxide was added to 10% H₂SO₃ (10 cm³) and set aside for 12 hours. The solution was diluted with H₂O and basified with NH₄OH and extracted with CHCl₃ (3 x 20 cm³). The extracts were combined, dried and solvent removed to afford a white powder m.p. 181° - 183°C; R_f 0.63 (EtOAc-Me₂CHOH-25% NH₄OH (9:7:2); UV: 278, 291, 305 nm. These data and TLC properties of the reduction product were identical with that of cocsuline.

Preparation of cocsuline N-oxide

2 mg of cocsuline was dissolved in 5 cm³ of MeOH and stirred with 15% H₂O₂ and left for 45 min at 25°C. The solution was diluted with a small amount of H₂O and extracted with CHCl₃. The CHCl₃ extract evaporated to afford a solid. The product was found to be a mixture of two compounds on TLC examination. The less polar of these had identical TLC properties and colour reactions as cocsuline N-oxide isolated from *Epinetrum villosum*.

Acknowledgements

The authors are grateful to Professors O.N. Ogbeide, F.C. Ohiri, Stermitz and Brochmann-Hanssen for authentic samples of alkaloids and ¹H-NMR spectra of cycleanine, cycleanine O-oxide and cocsuline.

References

1. F.R. Irvine, "Woody Plants of Ghana" Oxford University Press, London (1961).
2. J.O. Kokwaro, "Medicinal plants in East Africa" East African Literature Bureau, Nairobi (1976).
3. J.M. Watt and M.G. Breyer-Brandwijk, "The Medicinal and Poisonous Plants of South and Eastern Africa" E. & S. Livingstone Ltd., Edinburgh (1962).
4. A. Denoel, "Contribution a l etude chimique d'une Menispermacee congolaise: *Epinetrum villosum* (Exell) Troupin. Volume commemoratif du centenaire de L. Braemer, cehors, Impr. typo. A. Coueslant (1958).
5. A. Bouquet and A. Cave, *Plant Med. Phytother*, 5, 131 (1971).

6. F. Scheinmann, F. Scriven and O.N. Ogbeide *Phytochemistry*, **19**, 1835 (1980).
7. M. Debray, M. Plat and Le Men, *J. Ann. Pharm. Fr.* **24**, 551 (1966).
8. A.K. Bhtnagar, S. Bhattacharji and S.P. Popli, *Ind. J. Chem.*, **6**, 125 (1968).
9. D. Dwuma-Badu, J.S.K. Ayim, C.A. Mingle, A.N. Takie, D.J. Slatkin, J.E. Knapp and P.L. Schiff Jr., *Phytochemistry*, **14**, 2520 (1975).
10. T.Kikuchi and K.Bessho, *J. Pharm. Soc. (Japan)* **78**, 1408 (1958).
11. T. Kikuchi and K. Bessho, *J. Pharm. Soc. (Japan)*, **79**, 262 I(1959).
12. F.C. Ohiri, R. Verpoorte and A. Baerheim Svendsen, *Plant. Med. Phytother* **47**, 87 (1983).
13. D.S. Bhakuni and P.P. Joshi, *Tetrahedron*, **31**, 2575 (1975).
14. P.P. Joshi, D.S. Bhakuni and M.M. Dhar, *Ind. J. Chem.*, **12**, 649 (1974).
15. N. Bild and M. Hesse, *Helv. Chim. Acta*, **50**, 1887 (1967).
16. T.A. Bryce and J.R. Maxwell, *J. Chem. Soc., Chem. Commun.*, 206 (1965).
17. E. Weiss, K. Barnauer and A. Girardet, *Relv. Chim. Acta*, **54**, 1342 (1971).
18. J.D. Phillipson, and S.S. Handa, *Phytochemistry*, **14**, 2683 (1975).
19. H.H. Kondo and Tomita, *J. Pharm. Soc. (Japan)*, **52**, 139 (1932).
20. I.R.C. Bick and Todd, A.R. *J. Chem. Soc.*, 1606 (1950).
21. Silvestein - Bassler and Masrill: Spectrometric identification of organic compounds, ed., V John Wiley and Sons Inc., Singapore, Canada, p. 127 (1991).
22. D. Dwuma-Badu, D. Slatkin, J.E. Knapp and Schiff, Jr., *J. Pharm. Soc.*, **9**, 1242 (1977).
23. H. Guinaudeau N. Lin L.Z. Ruangrunsi and G.A. Cordell, *J. Natural Products*, **56**, 1989 (1993).