

Studies on Bioassay Directed Antifungal Activity of Medicinal Plants *Calotropis procera*, *Skimmia laureola*, *Peltophorum pterocarpum* and two pure Natural compounds uloptero and 4-methoxy-1-methyl-3-(2'S-hydroxy-3'-ene butyl)-2-quinolone

K.F. AHMAD* AND N. SULTANA
Pharmaceutical Research Centre,
PCSIR Laboratories Complex, Karachi, Pakistan

(Received 24th October, 2002, revised 28th July, 2003)

Summary: Our investigations on the antifungal activity of the crude ethanolic extracts of different plants including *Calotropis procera*, *Skimmia laureola* and *Peltophorum pterocarpum*, have led to the determination of good results of these activities. We report the evaluation of antifungal activity of crude extracts of *Calotropis procera*, *Skimmia laureola* and *P. pterocarpum*. In addition, we also report the antifungal activity of uloptero (1) and 4-methoxy-1-methyl-3-(2'S-hydroxy-3'-ene butyl)-2-quinolone (2) isolated from *Skimmia laureola* [1]. These plants were selected because of their reported medicinal importance in indigenous system of medicines from ancient times and are abundantly available in various parts of Pakistan [1,2].

Introduction

Calotropis procera (Ait) R. Br. (Asclepiadaceae), a xerophytic shrub widely distributed in the tropics of Asia and Africa is commonly known as "Akra" in India and Pakistan. In the traditional Indian medicinal system, different parts of the plant have been advocated for a variety of diseases and have also been considered as an antidote for snake poisoning [2]. It has been reputed in the Indian traditional medicine for a variety of ailments including leprosy, ulcer and piles. Different parts of the plant possess antimicrobial, anti-inflammatory, analgesic [3] and anticancer [4] activities.

Skimmia laureola is found in the Northern areas of Pakistan and is used in the indigenous system of medicine for the treatment of various ailments. The soot obtained from the burning of leaves is inhaled for treatment of body pain, fever and influenza [5].

Peltophorum pterocarpum is an ornamental tree grown in homes and gardens in India and Pakistan [6], bearing fragrant yellow flowers, reported to possess antibacterial, anti-inflammatory [7] and fungitoxic activity [8].

Results and Discussion

The crude extract of *Calotropis procera* exhibited antifungal activity [9] against *Trichophyton longiformis*, *Candida albicans*, *Aspergillus flavus*,

Microsporium canis, *Fusarium solani* and *Fusarium moniliformis*. The minimum inhibitory concentration (MIC) of crude extract against these fungi was used as 400 µg/ml. The growth of *Trichophyton longiformis*, *Candida albicans* and *Fusarium moniliformis* was inhibited in 100% by the crude extract at a concentration of 400 µg/ml, while standard fungicide Miconazole and Ketoconazole totally inhibited the growth at a concentration of 70 µg, 110.8 µg and 110.8 µg respectively. The growth of *Microsporium canis* was inhibited in 90% by the crude extract at a concentration of 400 µg/ml, while standard fungicide Miconazole totally inhibited the growth at the concentration of 98.4 µg/ml. These results are summarized in Table-1.

The crude ethanolic extract of *S. laureola* showed good activity against animal pathogen *Microsporium canis* and plant pathogen *Fusarium solani* var. *lycopersici* (Tomato) at a concentration of 400 µg/ml. Minimum inhibitory concentration (MIC) of the crude extract against *Microsporium canis* was used as 400 µg/ml. The growth of *Microsporium canis* was inhibited in 67.7% by the crude extract at a concentration of 400 µg/ml, while standard fungicide Miconazole and Ketoconazole totally inhibited the growth of *Microsporium canis* at a concentration of 72.10 and 62.25 µg/ml, respectively. The crude extract of *S. laureola* was found to be active against *Fusarium solani* var. *lycopersici* at a concentration of

*To whom all correspondence should be addressed.

Table-1: Antifungal activity of crude extract of *C. procera*.

| Fungi | Sample | Control | % Inhibition | Standard Drugs | % Inhibition | MIC $\mu\text{g/ml}$ |
|---------------------------------|--------|---------|--------------|-------------------------|--------------|----------------------|
| <i>Trichophyton longiformis</i> | 0 | 90 | 100 | Miconazole Ketoconazole | 100 | 70 |
| <i>Candida albicans</i> | 0 | 100 | 100 | Miconazole Ketoconazole | 100 | 110.8 |
| <i>Aspergillus flavus</i> | 100 | 100 | 0 | Amphotericin B | 100 | 70 |
| <i>Microsporium canis</i> | 10 | 100 | 90 | Miconazole Ketoconazole | 100 | 98.4 |
| <i>Fusarium solani</i> | 95 | 95 | 0 | Miconazole | 100 | 73.25 |
| <i>Fusarium moniliformis</i> | 0 | 100 | 100 | Miconazole | 100 | 110.8 |

Concentration of crude extract = 400 $\mu\text{g/ml}$.

Table-2 : Antifungal activity of crude extract of *Skimmia laureola*.

| Fungi | Crude extract of <i>S. laureola</i> | % Inhibition | Miconazole | Ketoconazole | Benlate | % Inhibition |
|---------------------------|-------------------------------------|--------------|------------|------------------------|------------------------|--------------|
| <i>Microsporium canis</i> | 400 $\mu\text{g/ml}$ | 67.7 | 72.10 | 62.25 $\mu\text{g/ml}$ | - | 100 |
| <i>Fusarium solani</i> | 400 $\mu\text{g/ml}$ | 57.7 | - | - | 73.25 $\mu\text{g/ml}$ | 100 |

Table-3 : Antifungal activity of ulopterol (1).

| Fungi | Compound (1) | % Inhibition | Miconazole | Ketoconazole | % Inhibition |
|----------------------------|----------------------|--------------|---------------------|---------------------|--------------|
| <i>Drechslera rostrata</i> | 200 $\mu\text{g/ml}$ | 50.8 | 25 $\mu\text{g/ml}$ | 25 $\mu\text{g/ml}$ | 100 |
| <i>Curvularia lunata</i> | 200 $\mu\text{g/ml}$ | 49.5 | 25 $\mu\text{g/ml}$ | 25 $\mu\text{g/ml}$ | 100 |

Table-4: Antifungal activity of 4-methoxy-1-methyl-3(2'S-hydroxy-3'-ene butyl)-2-quinolone (2).

| Fungi | Compound (2) | % Inhibition | Miconazole | Ketoconazole | % Inhibition |
|---------------------------------|----------------------|--------------|------------------------|------------------------|--------------|
| <i>Microsporium canis</i> | 200 $\mu\text{g/ml}$ | 68.7 | 72.10 $\mu\text{g/ml}$ | 62.25 $\mu\text{g/ml}$ | 100 |
| <i>Pseudoallescheria boydii</i> | 200 $\mu\text{g/ml}$ | 56.8 | 38.75 $\mu\text{g/ml}$ | 29.50 $\mu\text{g/ml}$ | 100 |

400 $\mu\text{g/ml}$, while standard fungicide Benlate completely inhibited the growth of *Fusarium solani* var. *lycopersici* at a concentration of 73.25 $\mu\text{g/ml}$. *Fusarium solani* var. *lycopersici* causes root rot, stem cankers associated with wounds, damping-off seedlings, destruction of spawn in beds of cultivated mushrooms and pea crop. The antifungal results are summarized in Table-2.

Compound (1) exhibited *in vitro* antifungal activity against *Drechslera rostrata* and *Curvularia lunata* with minimum inhibitory concentration of 200 $\mu\text{g/ml}$. Compound (1) also exhibited weak activity against *Curvularia lunata*. The fungicides Miconazole and Ketoconazole were used as standards. The results of antifungal assay results are summarized in Table-3.

Compound 2 exhibited some *in vitro* antifungal activity against *Microsporium canis* and *Pseudoallescheria boydii*. *Microsporium canis* causes infection of hair and skin in dogs and cats, while *Pseudoallescheria boydii* causes infection of skin, subcutaneous tissue, nasalsinuses and mycetoma. The minimum inhibitory concentration (MIC) of the compound 2, against *Microsporium canis* and *Pseudoallescheria boydii* was 200 $\mu\text{g/ml}$. The growth of *Microsporium canis* and *Pseudoallescheria boydii*

was strongly inhibited (68.7 and 56.8%) by the compound (2), while standard fungicide Miconazole and Ketoconazole totally inhibited the growth of *Microsporium canis* and *Pseudoallescheria boydii* at 72.10 $\mu\text{g/ml}$, 62.25 $\mu\text{g/ml}$ and 38.75 $\mu\text{g/ml}$, 29.50 $\mu\text{g/ml}$ concentration respectively [1,10]. The results of antifungal assay are summarized in Table-4.

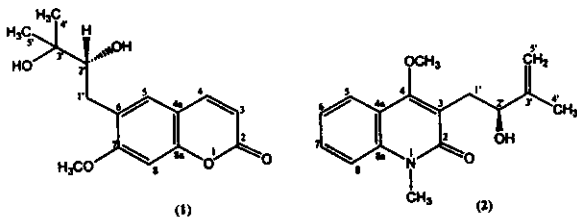
The crude extract of *P. pterocarpum* exhibited activity against a number of fungi. The *in vitro* antifungal activity was tested against *Trichophyton longifusus*, *Candida albicans*, *Aspergillus flavus*, *Microsporium canis*, *Fusarium solani* and *Candida glabrata*. Minimum inhibitory concentration of crude extract against *Microsporium canis* was used as 400 $\mu\text{g/ml}$. The growth of *Microsporium canis* was inhibited in 65% by the crude extract at a concentration of 400 $\mu\text{g/ml}$, while standard fungicide Miconazole totally inhibited the growth of *Microsporium canis* at a concentration of 98.4 $\mu\text{g/ml}$. The crude extract of *P. pterocarpum* was found to be active against *Trichophyton longifusus* and *Aspergillus flavus* at a concentration of 400 $\mu\text{g/ml}$, while standard fungicide Miconazole totally inhibited the growth of *Trichophyton longifusus* and *Aspergillus flavus* at a concentration of 70 $\mu\text{g/ml}$ and 20 $\mu\text{g/ml}$ respectively. The crude extract of *P. pterocarpum* exhibited no activity against *Candida*

Table-5: Antifungal activity of crude extract of *P. pterocarpum*

| Fungi | Sample | Control | % Inhibition | Standard drugs | MIC ($\mu\text{g/ml}$) |
|--------------------------------|--------|---------|--------------|----------------|--------------------------|
| <i>Trichophyton longifusus</i> | 70 | 95 | 26.3 | Miconazole | 70 |
| <i>Candida albicans</i> | 90 | 90 | 0 | Miconazole | 110.8 |
| <i>Aspergillus flavus</i> | 90 | 95 | 15.7 | Amphotericin B | 20 |
| <i>Microsporium canis</i> | 45 | 100 | 65 | Miconazole | 98.4 |
| <i>Fusarium solani</i> | 100 | 100 | 0 | Miconazole | 73.25 |
| <i>Candida glaberata</i> | 95 | 95 | 0 | Miconazole | 110.8 |

Concentration of sample = 400 $\mu\text{g/ml}$.

albicans, *Fusarium solani* and *Candida glaberata*. These results are summarized in Table-5.



Experimental

Calotropis procera, *Skimmia laureola* and *P. pterocarpum* were collected from the suburban areas of Karachi, Pakistan. The fresh dried plants of *C. procera* (1.0 kg), *S. laureola* (60 kg) and *P. pterocarpum* (1.0 kg) were ground and soaked in ethanol for 2 weeks and then filtered. The filtrate was concentrated under reduced pressure at 40°C to a gummy mass of *C. procera* (142.91 g), *S. laureola* (821.93 g) and *P. pterocarpum* (175.31 g). Ulopterol (1) and 4-methoxy-1-methyl-3-(2'S-hydroxy-3'-ene butyl)-2-quinolone (2) were isolated from alkaloidal fraction of *Skimmia laureola* by using extraction and column chromatography methods [1]. Antifungal screening has been performed on the crude extracts of *C. procera*, *S. laureola*, *P. pterocarpum* and two natural compounds 1 and 2. Sabouraud agar was used as antifungal control. Miconazole and Ketoconazole were used as standard drug.

Antifungal Bioassay

All antifungal assay employed a standard Agar Tube Dilution Method [11]. The test fungi were maintained on Sabouraud's agar slants. A 4 mm diameter piece of fungal inoculum removed from 7 days old culture of fungi was transformed in solid media. The test sample of crude extract was dissolved in sterile DMSO to obtain 200 $\mu\text{g/ml}$ concentration. Sabouraud agar was prepared by mixing 32.5 gram Sabouraud dextrose agar, 4% glucose agar and 7.5 gram of agar-agar in 500 ml distilled water. Tubes were allowed to cool to 50°C and non-solidified

Sabouraud agar media were inoculated with 200 μl of compound pipetted from the stock solution. This gave a final concentration of 200 $\mu\text{g/ml}$ of media. The tubes were then allowed to solidify in a slanted position at room temperature and the tubes were incubated at 27-29°C for 7-10 days and the visible growth observed. Minimum inhibitory concentrations (MIC) were expressed in $\mu\text{g/ml}$.

Acknowledgment

The authors fully acknowledge and highly appreciate the cooperation, support and kind help of Professor Dr. Muhammad Iqbal Choudhary for allowing us to conduct Bioassay studies of these medicinal plants at HEJ Research Institute of Chemistry, University of Karachi.

References

1. N. Sultana, Phytochemical and structural studies on the chemical constituents of *Adhatoda vasica*, *Sarcococca salignu* and *Skimmia laureola*, Ph.D. Dissertation, HEJ Research Institute of Chemistry, University of Karachi, Karachi-75270, Pakistan, (2000).
2. A.K. Nandkarni, *Indian Materia Medica*, Popular Book Depot, Bombay, 1, 246 (1976).
3. A. Basu and A.K. Nag Chaudhuri, *Journal of Ethnopharmacology*, 31, 319 (1991).
4. S.M. Hussein Ayoub and D.G.I. Kingston, *Fitoterapia*, 52, 281 (1981).
5. K.M. Nadkarni, *Indian Materia Medica*, Popular Prakashan, Bombay, 1, 1142 (1976).
6. J.D. Hooker, *The Flora of British India*, Reeve and Co., England, 2, 257 (1954).
7. R. Sethu, M.G. Sulochna and N. Lalithakameswari, *Fitoterapia*, IV(3), 177 (1984).
8. S. Rao, R. Seeta and K.V.N. Rao, *Indian Journal of Plant Physiol.*, 29(3), 278 (1986).
9. J.F. Couch, *J. Am. Chem. Soc.*, 59, 1469 (1937).
10. Atta-ur-Rahman, N. Sultana, M.I. Choudhary, P.M. Shah and M.R. Khan, *J. Nat. Prod.*, 61, 713 (1998).
11. C. Leben and G.W. Keit, *Phytopathology*, 32, 814 (1947).