

Determination of the Binding Constants, Bound Chemical Shifts, And Stoichiometry of Lanthanide-Substrate Complexes

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(Received 1st December, 1982)

Summary: Analysis of the experimental results of the equilibrium between a lanthanide shift reagent, L, and a substrate, S, is presented. When different methods were used, evaluation of equilibrium binding constants (K_B), bound chemical shift (Δ_B), and stoichiometry (n) of lanthanide (L)-substrate (S) complexes brought about the same numerical values under the conditions of $[S]_0 \gg [L]_0$. Finally, it is shown that the association between the lanthanide and the substrate (i.e. 1-(X-benzo [b] thienyl- ethyl acetate derivatives) has 1:1 stoichiometry.

Introduction

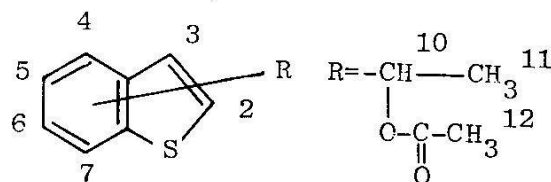
Lanthanide shift reagent (LSR'S) have proved to be of considerable value for evaluation of molecular structure using nuclear magnetic resonance, both as a qualitative aid in simplifying spectra and as a quantitative means of correlating molecular structures by means of the pseudocontact equation¹. In this paper we present experimental data for several compounds which demonstrate that the different methods lead to the same value K_B binding constants, Δ_B bound chemical shifts and the stoichiometry of lanthanide-substrate complexes were also determined.

Addition of Eu (FOD)₃ to the chloroform solution of benzo [b] thiophene under the same condition used for (benzo [b] thienyl) ethyl acetate derivatives show no appreciable proton shifts, but only broad peaks were observed. This indicates that the induced shifts are not consistent with complexing involving the sulphur atom; on the other hand, addition of Eu (FOD)₃ to the (benzo [b] - thienyl) ethyl acetate derivatives was very useful. Ester group is the only site of substrate able to react with the induced shift reagents. Earlier results confirmed that thioamides involve the sulphur atom, when forming complexes with shift reagents^{2,3}. But sulfoxides and sulfones complex readily via the oxygen atom⁴.

Results and Discussion

Esters are weaker Lewis bases than ketones toward

LSR⁵, the preferred coordination site being the carbonyl oxygen. For this reason, the stronger Lewis acid Eu (FOD)₃ was initially employed for the simplification of the spectra of simple esters⁶. Analysis of the chemical shifts of the rings and the ring side chain protons spectra after the addition of Eu (FOD)₃ to compounds 1-(X-benzo [b] thienyl) ethyl acetate derivatives, X = 2 to 7 are listed in Table 1 with the induced downfield shifts.



Observed chemical shifts of the ring side chain protons (H_{10} , H_{11} & H_{12}) illustrate that H_{10} is strongly affected by LSR than H_{12} while H_{11} is less affected by LSR than H_{12} . The same results were obtained by addition of Eu (FOD)₃ to 1-phenylethyl acetate and 1-phenylethyl alcohol. This may indicate that the oxygen atom of the ester group is the favoured binding site than the carbonyl oxygen⁷.

Much progress has been made in applying LSR's to the study of molecular geometry in solution and excellent techniques for this are now available⁸. However, structural determinations have nearly all been derived from observed lanthanide induced shifts (LIS)

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Table 1. Lanthanide-Induced Downfield Shifts (Hz) in ^1H NMR Spectra of 1-(X-Benzo [b] thienylethyl Acetate Derivatives^a.

Compound	Substituent		H ₂	H ₃	H ₄	H ₅	H ₆	H ₇	H ₁₀	H ₁₁	H ₁₂
2	2-R	δ	—	8.38	7.90	7.45	7.41	8.09	10.2	3.03	5.04
		$\Delta\delta$	—	115	22	16	14	33	400	136	297
3	3-R		8.46	—	9.24	7.68	7.56	8.19	10.5	3.21	5.1
			107	—	144	34	26	35	419	153	303
4	4-R		7.66	8.79	—	8.52	7.71	8.17	10.23	2.97	4.74
			20	125	—	115	42	36	389	133	267
5	5-R		7.58	7.52	9.00	—	8.57	8.15	9.98	2.95	4.93
			20	28	128	—	133	35	398	139	289
6	6-R		7.58	7.47	8.14	8.51	—	9.05	9.75	2.87	4.83
			18	19	39	120	—	119	375	128	275
7	7-R		7.80	7.71	8.18	7.84	8.61	—	10.44	3.13	5.08
			38	33	45	45	135	—	428	146	296
8	1-phenylethyl-acetate		<u>Ho</u>	<u>Hm</u>	<u>Hp</u>	—	—	—	11.61	3.50	6.27
			9.12	7.93	7.85	—	—	—	—	—	—
			183	64	56	—	—	—	574	205	426
9	1-phenylethyl-alcohol		<u>Ho</u>	<u>Hm</u>	<u>Hp</u>	—	—	—	10.86	4.88	<u>-OH</u>
			10.30	8.21	8.02	—	—	—	—	—	7.26
			296	87	68	—	—	—	579	339	541

^a = In chloroform-d 0.1M solution. $\Delta\delta$ = is the differences in Hz before and after the addition of $\text{Eu}(\text{FOD})_3$.

rather than from the intrinsic parameters, the limiting incremental shifts. Also the equilibrium constants are of some importance since they give information on the stability of a complex. It has been reported by Armitage et al⁹, and of Kelsey¹⁰ that the LIS obey an equation of the following form, under the conditions $S_0 \gg L_0$ (see experimental method 1).

$$[S]_0 = [L]_0 \Delta_B (1/\Delta\delta) - ((1/K_B) + [L]_0) \dots 1$$

Thus equation 1 gives the important fact that a plot of $[S]_0$ vs $1/\Delta\delta$ (at constant L_0) gives a straight line whose slope is $[L]_0 \Delta_B$, and whose Y-intercept is $-(1/K_B) + [L]_0$. Such a plot thus yield both Δ_B and K_B unambiguously. This equation was derived under the assumption of a 1:1 complex formation

and is very similar to the Scott¹¹ modification of the Benesi-Hilderbrand equation¹².

The fits^{9,10} so far obtained to eq. 1 have yielded excellent straight lines and this has been used as evidence for the simple one-step mechanism. The results are shown in Fig. 1. Here are plotted the substrate concentration $[S]_0$ of 1-phenylethyl acetate vs $1/\Delta\delta$ 1-phenylethyl acetate. This compound was chosen here because of the fact that the protons are not obscured by fused ring protons as in (benzo [b] thienyl) ethyl acetate derivatives. The values of K_B and Δ_B were obtained from these plots following the relationship given in eq. 1; the numerical values for these parameters are shown in Table 2. It is expected that the same value should be obtained for K_B regardless of which proton is used for its determination. This may be confirmed

Table 2. Calculated Values of Bound Chemical Shifts (Δ_B), Binding Constants (K_B) and Stoichiometry for Complexes of Organic Substrates with Eu (FOD)₃.

Substrate	Δ_B^*		K_B^\ddagger	Stoichiometry		
	(1)	(2)		(2)	(3)	
1-phenylethyl- acetate	H ₁₀	8.4	8.8	≥ 100	1.2	0.7
	H ₁₁	2.9	3.1	≥ 100	1.1	
	H ₁₂	5.9	6.5	≥ 100	1.1	
1-(4-Benzo[b]- thienyl-ethyl- acetate	H ₁₀	6.0		≥ 100	1.1	
	H ₁₁	2.0		≥ 100	1.1	
	H ₁₂	4.5		≥ 100	1.1	

*Values of Δ_B (ppm) and K_B (1 mol⁻¹) derived from method (1) and method(2).

(1) Method 1, (see experimental)

(2) Method 2, (see experimental)

(3) This value should be regarded with some skepticism (See text and eq.2.).

experimentally by noting that the plots for protons H₁₀, H₁₁ and H₁₂ in Fig. 1, all intercept the Y-axis at the same point. It is worthnoting that the Δ_B values obtained by this method vary inversely with distance from the coordination site, as expected for a pseudo-contact shift. K_B was too large to be measured, the Y-intercept depends on (1/ K_B), and when K_B is large, the Y-intercept is so close to zero that K_B cannot be determined with any confidence; in this case only a lower limit for K_B may be deduced. It is well known that Eu (FOD)₃ form a strong binding complex with the substrate, compared with Eu (DPM)₃ complex⁹.

The calculated Δ_B are consistent with Δ_B i/ Δ_{BJ} slopes, e.g., Δ_B for H₁₀ and H₁₂ are calculated to be 8.4 and 5.9 ppm⁻¹; ratio 1.4. The plot of Δ_{Obsd} H₁₀ vs Δ_{Obsd} H₁₂ gives slope 1.4 (see Fig. 2). This analysis allows convenient estimations of Δ_B and K_B .

In other experiments (method 2), in which [L]₀ is varied at constant [S]₀ and $S_0 \gg L_0$, for strong binding or high substrate concentration, a derived equation was obtained⁹.

$$\delta = \Delta_B [L]_0 / [S]_0 \dots\dots\dots 2$$

Thus plots of δ vs [L]₀ / [S]₀ , gives a slope of

Δ_B . Fig. 3 illustrates these observed shifts and Fig. 4 shows the same shifts as a function of the sum of the observed shifts of all the signals in the spectrum. Clearly, the plotting of shift vs shift has the effect of linearizing the data. The numerical values of Δ_B obtained from Fig. 3 are nearly the same of that obtained by method 1 (see Table 2).

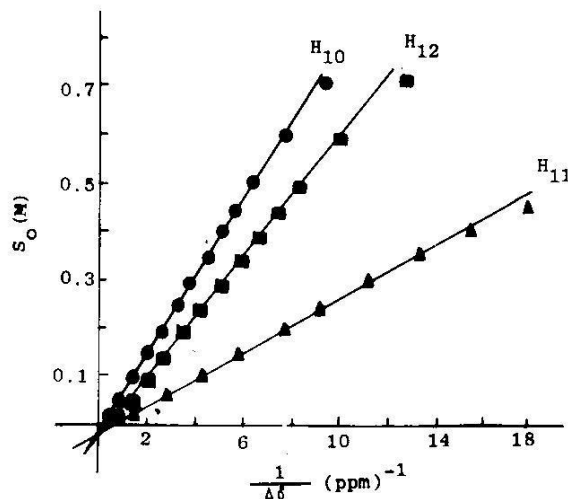


Fig.1

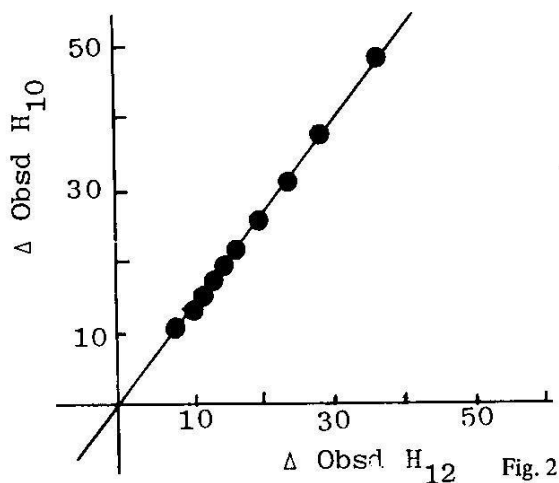


Fig. 2

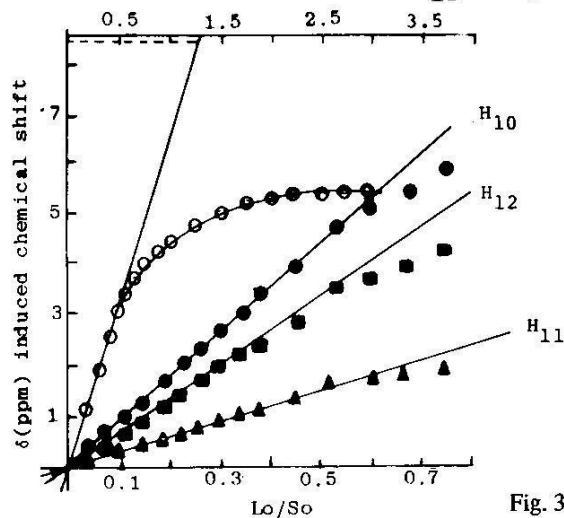


Fig. 3

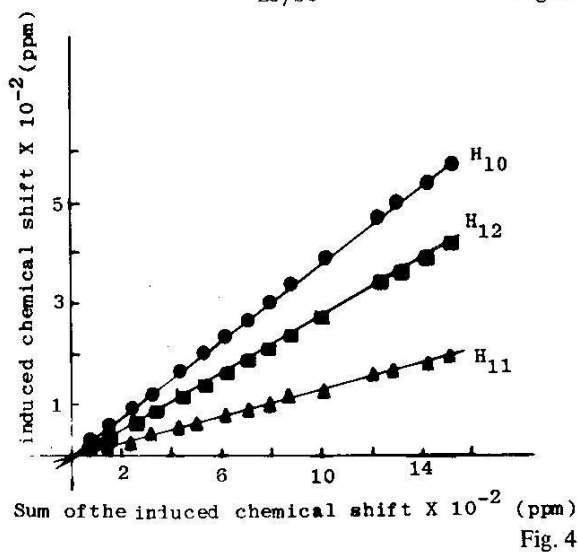


Fig. 4

Stoichiometry of the Complex

The stoichiometry of the complex can be estimated graphically from the position of maximum curvature of the $[L]_0 / [S]_0$ axis of a shift diagram⁹ (Fig. 3). In this case, the limiting line (the left-most points in Fig. 3) extrapolated to the known $\Delta\beta$ value. The stoichiometry appears to be 1:1. Further evidence for the stoichiometry of 1:1 gained from the straight lines in Fig. 1 (see text).

More precise method have been described for the determination of the stoichiometry n , using eq. 3¹³.

$$\log [S] = 1/n [\log ([LS] / [L]) - \log K] \dots 3$$

Thus a plot of $\log [S]$ vs $\log ([LS] / [L])$ will have a slope $1/n$ and $\log K$ from intercept, giving a direct measurement of the stoichiometry of the complex (see Table 2). However, for large values of K , eq. 2 gives uncertain values of n ¹³.

Experimental:

All NMR spectra were measured on JEOL JNM FX-100 spectrometer operating at 100 MHz in the Fourier Transform Mode. All the spectra were recorded at ambient temperature 25^o and over 1000 and 1500 Hz sweep width using 16 K data points. Chemical shifts are in δ units (part per million) from internal TMS. Heterocyclic compounds of the title were prepared by known methods^{14,15}. The LSR used in these studies was Europium (III) - tris- (1,1,1,2,2,3,3,- heptafluoro-7, 7-dimethyl-4, 6-octadionate). Eu (FOD)₃, was supplied by FLUKA AG. The solutions of shift reagent were prepared freshly prior to use but generally no further precautions were taken to exclude moisture.

The NMR runs for data were fit to Fig. 1 to 4 were performed in the following manner:

Method 1: Experiments in which $[S]_0$ is varied at constant $[L]_0$.

The lanthanide reagent (5.3 mg, 0.01 M) of Eu (FOD)₃ was dissolved in chloroform-d (0.5 ml), in a clean, oven-dried NMR tube with 0.01 ml TMS. The substrate was injected into the NMR tube with thorough mixing through the plastic cap using a syringe graduated in 1 μ l increments. The NMR tube was inserted to the NMR probe and scan was done. More substrate was added as before, and scan were repeated until

excess of substrate 60 μ l were added (ca. 0.7 M) and spectra were recorded for a total of 13 different concentrations of substrate (Data were fit to Fig. 1 and 2).

Method: 2 Experiments in which $[L]_0$ is varied at constant $[S]_0$:

(A) 0.5 M of substrate in chloroform-d (0.5 ml) was prepared in a clean, oven-dried NMR tube with 0.01 ml TMS. The NMR tube was inserted to the NMR probe and first scan was recorded. The lanthanide reagent of 10 mg increments were added with thorough mixing to the NMR tube and scans were repeated until a total of 200 mg were added (15 different concentrations of lanthanide reagents), data were fit to Fig. 3 and 4).

(B) The initial sample was prepared in a clean, oven-dried NMR tube by first putting in \sim 155 mg of Eu (FOD)₃ (0.3 M) in chloroform-d solution (0.5 ml) with 8.2 mg (0.1 M) of substrate in the NMR tube. The original spectrum was recorded. The incremental dilution method¹⁶ was used, in which the initial sample was successively diluted with a 0.1 M chloroform-d solution of the substrate. Thus the concentration of the substrate remains constant at 0.1 M while the concentration of the shift reagent decreases with each dilution. Spectra were recorded for a total of 25 different concentrations of shift reagent (data were fit to Fig. 3, the broken curve).

Acknowledgements:

The authors gratefully acknowledge the technical assistance of Mr. R.M. Saad. We also thank the Research

Centre, College of Science, King Saud University (formerly Riyadh University) for financial support.

References:

1. D.J. Raber, M.D. Johnston, C.M. Campbell, C.M. Tanks and P. Sutton, *Org. Magn. Reson.*, 11, 323 (1978) and references cited therein.
2. W. Walter, R.F. Becker and J. Thiem, *Tetrahedron Lett.*, 1971 (1971).
3. A.H. Lewin, *Tetrahedron Lett.*, 3583 (1971).
4. K.K. Anderson and J.J. Uebel, *Tetrahedron Lett.*, 5253 (1971).
5. J.K.M. Sanders and D.H. Williams, *J. Amer. Chem. Soc.*, 93, 641 (1971).
6. R.G. Rondeau and R.E. Sievers, *J. Amer. Chem. Soc.*, 93 1522 (1971).
7. H.B. Amin, S.S. Al-Showiman and I.M. Al-Najjar, *J. Chem. Soc. Pak.* 4 (3), 155, (1982).
8. A.F. Cockerill, G.L. Odavies, R.C. Harden and D.M. Rackham, *Chem. Rev.* 73 (6) 553, (1973).
9. I. Armitage, G. Dunsmore, L.D. Hall and A.G. Marshall, *Canad. J. Chem.*, 50, 2119 (1972).
10. D.R. Kelsey, *J. Amer. Chem. Soc.*, 94, 1764 (1972).
11. R.L. Scott, *ibid.*, 75, 787 (1956).
12. H.A. Benesi and J.H. Hildebrand, *ibid.*, 71, 2703 (1949).
13. I. Armitage, G. Dunsmore, L.D. Hall and A.G. Marshall, *Chem. Ind. (London)*, 79 (1972).
14. R. Taylor, *J. Chem. Soc. (B)*, 2382 (1971).
15. H.B. Amin and R. Taylor, *J. Chem. Soc. Perkin II*, 1053, (1978).